

**Mouse CSF1R/M-CSFR ELISA Kit**  
**(mCSF1R-ELISA)***Cat. No. EK0808**96 Tests in 8 x 12 divisible strips***Background**

CSF1R (Colony stimulating factor 1 receptor), also known as M-CSFR and CD115, is a cell-surface protein encoded, in humans, by the CSF1R gene. The gene is located on the long arm of chromosome 5 (5q32) on the Crick (minus) strand. The encoded protein is a tyrosine kinase transmembrane receptor and a member of the CSF1/PDGF receptor family of tyrosine-protein kinases. The encoded protein is a single pass type I membrane protein and acts as the receptor for colony stimulating factor 1 (CSF1), a cytokine which controls the production, differentiation, and function of macrophages. Both CSF1R and its ligand CSF1 play an important role in the development of the mammary gland and may be involved in the process of mammary gland carcinogenesis.

ScienCell's mouse CSF1R ELISA kit is based on standard sandwich enzyme-linked immunosorbent assay technology. Mouse CSF1R-specific polyclonal antibodies are pre-coated onto 8 x 12 divisible strips. The mouse specific detection monoclonal antibodies are biotinylated. The test samples and biotinylated detection antibodies are subsequently added to the wells and then washed with PBS or TBS buffer. Avidin-Biotin-Peroxidase Complex is added and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB is used to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product that changes to yellow after adding acidic stop solution. The intensity of yellow is proportional to the amount of mouse CSF1R in the sample that is captured on the strips.

<b>Size</b>	96 Tests in 8 x 12 divisible strips
<b>Assay type</b>	Sandwich ELISA
<b>Range</b>	62.5 pg/ml-4000 pg/ml
<b>Sensitivity</b>	< 10 pg/ml
<b>Specificity</b>	Natural and recombinant mouse CSF1R
<b>Cross-reactivity</b>	No detectable cross-reactivity with other relevant proteins.
<b>Storage</b>	Store at 4°C for frequent use, at -20°C for infrequent use. Avoid multiple freeze-thaw cycles.
<b>Shipping</b>	Shipped with gel ice.
<b>Expiration</b>	Four months at 4°C and eight months at -20°C.

<b>Application</b>	For quantitative detection of mouse CSF1R in cell culture supernatants, serum and plasma (heparin, EDTA).
<b>Kit components</b>	<ol style="list-style-type: none"> <li>1. Lyophilized recombinant mouse CSF1R standard: 10ng/tube×2.</li> <li>2. 8 x 12 divisible strips pre-coated with anti- mouse CSF1R antibody.</li> <li>3. Sample diluent buffer: 30 ml</li> <li>4. Biotinylated anti-mouse CSF1R antibody: 130µl, dilution 1:100.</li> <li>5. Antibody diluent buffer: 12ml.</li> <li>6. Avidin-Biotin-Peroxidase Complex (ABC): 130µl, dilution 1:100.</li> <li>7. TMB color developing agent: 10ml.</li> <li>8. TMB stop solution: 10ml.</li> </ol>
<b>Materials Required But Not Provided</b>	<ol style="list-style-type: none"> <li>1. Microplate reader.</li> <li>2. Automated plate washer.</li> <li>3. Adjustable pipettes and pipette tips. Multichannel pipettes are recommended for large amount of samples.</li> <li>4. Clean tubes and Eppendorf tubes.</li> <li>5. Washing buffer (neutral PBS or TBS).</li> </ol> <p>Preparation of 0.01M TBS: Add 1.2g Tris, 8.5g NaCl; 450µl of purified acetic acid or 700µl of concentrated hydrochloric acid to 1000ml H<sub>2</sub>O and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.</p> <p>Preparation of 0.01 M PBS: Add 8.5g NaCl, 1.4g Na<sub>2</sub>HPO<sub>4</sub> and 0.2g NaH<sub>2</sub>PO<sub>4</sub> to 1000ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.</p>
<b>Usage</b>	This product is for research use only. It is not approved for use in humans, animals, or <i>in vitro</i> diagnostic procedures.

## Reference

1. Galland F, Stefanova M, Lafage M, Birnbaum D (1992). "Localization of the 5' end of the MCF2 oncogene to human chromosome 15q15→q23". Cytogenet. Cell Genet. 60 (2): 114–6.
2. Tamimi RM, Brugge JS, Freedman ML, Miron A, Iglehart JD, Colditz GA, Hankinson SE (January 2008).
3. Pollard JW, Hennighausen L (September 1994). "Colony stimulating factor 1 is required for mammary gland development during pregnancy". Proc. Natl. Acad. Sci. U.S.A. 91 (20): 9312–6.
4. Sapi E (January 2004). "The role of CSF-1 in normal physiology of mammary gland and breast cancer: an update". Exp. Biol. Med. (Maywood) 229 (1): 1–11.

## Protocol for Mouse CSF1R ELISA (96-well format)

### Notes before you begin

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, a pilot experiment using standards and a small number of samples is recommended.
2. The TMB Color developing agent should be colorless and transparent before using.
3. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
4. A duplicate well assay is recommended for both standard and samples.
5. Do not let wells dry, as this will inactivate active components in wells.
6. Do not reuse tips and tubes to avoid cross contamination.

7. Avoid using reagents from different batches.
8. In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37°C for 30 minutes before use.

## **Preparation**

### **Sample Preparation and Storage**

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C.

Avoid repeated freeze-thaw cycles.

- **Cell culture supernatants:** Remove particulates by centrifugation, assay immediately or aliquot and store at -20°C.
- **Serum:** Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 minutes. Analyze the serum immediately or aliquot and store samples at -20°C.
- **Plasma:** Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 minutes at 1500 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C.

### **Sample Dilution Guideline**

The user needs to estimate the concentration of the target protein in the sample and select a proper dilution factor so that the diluted target protein concentration falls near the middle of the linear regime in the standard curve. Dilute the sample using the provided diluent buffer. The following is a guideline for sample dilution. Several trials may be necessary in practice. **The sample must be mixed well with the diluent buffer.**

- **High target protein concentration (40-400 ng/ml).** The working dilution is 1:100. i.e. Add 3 µl sample into 297 µl sample diluent buffer.
- **Medium target protein concentration (4-40 ng/ml).** The working dilution is 1:10. i.e. Add 25 µl sample into 225 µl sample diluent buffer.
- **Low target protein concentration (62.5-4000 pg/ml).** The working dilution is 1:2. i.e. Add 100 µl sample to 100 µl sample diluent buffer.
- **Very Low target protein concentration (≤62.5 pg/ml).** No dilution necessary, or the working dilution is 1:2.

### **Reagent Preparation and Storage**

A. Reconstitution of the mouse CSF1R standard: CSF1R standard solution should be prepared no more than 2 hours prior to the experiment. Two tubes of CSF1R standard (10 ng per tube) are included in each kit. Use one tube for each experiment.

- 10,000 pg/ml of mouse CSF1R standard solution: Add 1 ml sample diluent buffer into one tube, keep the tube at room temperature for 10 minutes and mix thoroughly.
- 4000pg/ml of mouse CSF1R standard solution: Add 0.4ml of the above 10ng/ml CSF1R standard solution into 0.6ml sample diluent buffer and mix thoroughly.
- 2000pg/ml→62.5pg/ml of mouse CSF1R standard solutions: Label 6 Eppendorf tubes with 2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml respectively. Aliquot 0.3ml of the sample diluent buffer into each tube. Add 0.3ml of the above 4000pg/ml CSF1R standard solution into 1st tube and mix. Transfer 0.3 ml from 1st tube to 2nd tube and mix. Transfer 0.3ml from 2nd tube to 3rd tube and mix, and so on.

**Note:** The standard solutions are best used within 2 hours. The 10 ng/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

B. Preparation of biotinylated anti-mouse CSF1R antibody working solution: The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).
  - Biotinylated anti-mouse CSF1R antibody should be diluted in 1:100 with the antibody diluent buffer and mixed thoroughly (i.e. Add 1µl Biotinylated anti-mouse CSF1R antibody to 99µl antibody diluent buffer).
- C. Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution: The solution should be prepared no more than 1 hour prior to the experiment.
- The total volume should be: 0.1ml/well x (the number of wells). (Allowing 0.1-0.2 ml more than total volume).
  - Avidin-Biotin-Peroxidase Complex (ABC) should be diluted in 1:100 with the ABC dilution buffer and mixed thoroughly (i.e. Add 1µl ABC to 99µl ABC diluent buffer).

### Assay Procedure

The ABC working solution and TMB color developing agent must be kept warm at 37°C for 30 minutes before use. When diluting samples and reagents, they must be mixed completely and evenly. Standard CSF1R detection curve should be prepared for each experiment. The user will decide sample dilution fold by crude estimation of CSF1R amount in samples.

1. Aliquot 0.1ml per well of the 4000pg/ml, 2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml mouse CSF1R standard solutions into the pre-coated 8 x 12 divisible strips. Add 0.1ml of the sample diluent buffer into the control well (**blank well**). Add 0.1ml of each properly diluted sample of mouse cell culture supernatants, serum or plasma (heparin, EDTA) to each empty well. See “**Sample Dilution Guideline**” above for details. We recommend that each mouse CSF1R standard solution and each sample is measured in duplicate.
2. Seal the strips with the cover and incubate at 37°C for 90 minutes.
3. Remove the cover, discard the strips’ contents, and blot the strips onto paper towels or other absorbent material. **Do NOT** let the wells completely dry at any time.
4. Add 0.1ml of biotinylated anti-mouse CSF1R antibody working solution into each well and incubate the strips at 37°C for 60 minutes.
5. Wash the strips 3 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1 minute. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (**Strips Washing Method**: Discard the solution in the wells without touching the side walls. Blot the strips onto paper towels or other absorbent material. Soak each well with at least 0.3 ml PBS or TBS buffer for 1~2 minutes. Repeat this process two additional times for a total of THREE washes. Note: For automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling wells with PBS or TBS buffer. Blot the strips onto paper towels or other absorbent material).
6. Add 0.1ml of prepared ABC working solution into each well and incubate the strips at 37°C for 30 minutes.
7. Wash the strips 5 times with 0.01M TBS or 0.01M PBS, and each time let washing buffer stay in the wells for 1-2 minutes. Discard the washing buffer and blot the strips onto paper towels or other absorbent material. (See Step 5 for strip washing method).
8. Add 90 µl of prepared TMB color developing agent into each well and incubate the strips at 37°C in dark for 20-25 minutes (**Note**: For reference only, the optimal incubation time should be determined by end user. And the shades of blue can be seen in the wells with the four most concentrated mouse CSF1R standard solutions; the other wells show no obvious color).
9. Add 0.1ml of prepared TMB stop solution into each well. The color changes to yellow immediately.
10. Read the O.D. absorbance at 450 nm in a microplate reader within 30 minutes after adding the stop solution.

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of blank well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The mouse CSF1R concentration of the samples can be interpolated from the standard curve.

**Note:** if the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

### **Summary**

1. Add samples and standards and incubate the strips at 37°C for 90 minutes. Do not wash.
2. Add biotinylated antibodies and incubate the strips at 37°C for 60 minutes. Wash strips 3 times with 0.01M TBS.
3. Add ABC working solution and incubate the strips at 37°C for 30 minutes. Wash strips 5 times with 0.01M TBS.
4. Add TMB color developing agent and incubate the strips at 37°C in dark for 20-25 minutes.
5. Add TMB stop solution and read.

### **Typical Data Obtained from Mouse CSF1R**

(TMB reaction incubate at 37°C for 20 minutes)

Concentration (pg/ml)	0.0	62.5	125	250	500	1000	2000	4000
Absorbance (450 nm)	0.034	0.078	0.135	0.198	0.354	0.706	1.106	2.093

### **Typical Mouse CSF1R ELISA Kit Standard Curve**

This standard curve was generated for demonstration purpose only. A standard curve must be run with each assay.

